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**Term:**

L1 same (acidothermus cellulolyticus)

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DB=USPT,PGPB,JPAB,EPAB,DWPI; PLUR=YES; OP=ADJ

<u>L7</u>	L5 same exoglucanase	2	<u>L7</u>
<u>L6</u>	L5 same (family 48)	0	<u>L6</u>
<u>L5</u>	L1 same (acidothermus cellulolyticus)	22	<u>L5</u>
<u>L4</u>	L1 same (glycoside hydrolase)	38	<u>L4</u>
<u>L3</u>	L1 same (GH48 or family 48 or Gux1)	0	<u>L3</u>
<u>L2</u>	L1 same (GH48 or family 48 or Gux1)	0	<u>L2</u>
<u>L1</u>	cellulase	8139	<u>L1</u>

END OF SEARCH HISTORY

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L5: Entry 7 of 22

File: USPT

Nov 9, 1999

DOCUMENT-IDENTIFIER: US 5981835 A

TITLE: Transgenic plants as an alternative source of lignocellulosic-degrading enzymes

Detailed Description Text (243):

Tucker et al. (1989), Ultra-thermostable cellulases from Acidothermus cellulolyticus comparison of temperature optima with previously reported cellulases. Biotechnology 7:817-820.

Other Reference Publication (30):

Tucker et al. (1989), Ultra-thermostable cellulases from Acidothermus cellulolyticus comparison of temperature optima with previously reported cellulases. Biotechnology 7:817-820.

=> d his

(FILE 'HOME' ENTERED AT 08:40:02 ON 12 FEB 2003)

INDEX 'ADISCTI, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI, BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO, CABA, CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DDFB, DDFU, DGENE, DRUGB, DRUGLAUNCH, DRUGMONOG2, ...' ENTERED AT 08:40:13 ON 12 FEB 2003

SEA (GLYCOSIDE HYDROLASE) OR (GLYCOSYL HYDROLASE)

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1326 FILE AGRICOLA  
34 FILE AQUASCI  
18 FILE BIOBUSINESS  
3 FILE BIOCOMMERCE  
701 FILE BIOSIS  
128 FILE BIOTECHABS  
128 FILE BIOTECHDS  
468 FILE BIOTECHNO  
2766 FILE CABA  
699 FILE CANCERLIT  
841 FILE CAPLUS  
29 FILE CEABA-VTB  
12 FILE CONFSCI  
1 FILE CROPB  
1 FILE CROPU  
4909 FILE DDFB  
3 FILE DDFU  
384 FILE DGENE  
4909 FILE DRUGB  
7 FILE DRUGU  
18 FILE EMBAL  
496 FILE EMBASE  
533 FILE ESBIODBASE  
16 FILE FEDRIP  
18 FILE FROSTI  
205 FILE FSTA  
509 FILE GENBANK  
29 FILE IFIPAT  
16747 FILE JICST-EPLUS  
354 FILE LIFESCI  
10631 FILE MEDLINE  
98 FILE NTIS  
7 FILE OCEAN  
215 FILE PASCAL  
1 FILE PHAR  
2 FILE PROMT  
968 FILE SCISEARCH  
1170 FILE TOXCENTER  
292 FILE USPATFULL  
5 FILE USPAT2  
169 FILE VETB  
56 FILE WPIDS  
56 FILE WPINDEX

L1 QUE (GLYCOSIDE HYDROLASE) OR (GLYCOSYL HYDROLASE)

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FILE 'JICST-EPLUS, MEDLINE, DRUGB, CABA, AGRICOLA, TOXCENTER, SCISEARCH, CAPLUS, BIOSIS' ENTERED AT 08:42:11 ON 12 FEB 2003

L2 0 S L1 AND (GH48) OR (GUX1)  
L3 51 S L1 AND (FAMILY 48)  
L4 4 S L3 AND (ACIDOTHERMUS OR CELLULOLYTICUS)  
L5 0 S L1 AND (GH48 OR GUX1)

L6           1 DUP REM L4 (3 DUPLICATES REMOVED)  
L7           13 DUP REM L3 (38 DUPLICATES REMOVED)  
L8           45712 S CELLULASE  
L9           49 S L8 AND (FAMILY 48 OR GH48)  
L10          110 S L8 AND (ACIDOTHERMUS CELLULOLYTICUS)  
L11          0 S L10 AND (FAMILY 48 OR GH48 OR GUX1)  
L12          16 S L10 AND (EXOGLUCANASE OR CBD)  
L13          4 DUP REM L12 (12 DUPLICATES REMOVED)  
L14          17 DUP REM L9 (32 DUPLICATES REMOVED)

=> d 113 ibib ab 1-4

L13 ANSWER 1 OF 4 MEDLINE DUPLICATE 1  
ACCESSION NUMBER: 1998290861 MEDLINE  
DOCUMENT NUMBER: 98290861 PubMed ID: 9627391  
TITLE: Hydrolysis of cellulose using ternary mixtures of purified  
**cellulases**.  
AUTHOR: Baker J O; Ehrman C I; Adney W S; Thomas S R; Himmel M E  
CORPORATE SOURCE: Biotechnology Center for Fuels and Chemicals, National  
Renewable Energy Laboratory, Golden, CO 8040, USA.  
SOURCE: APPLIED BIOCHEMISTRY AND BIOTECHNOLOGY, (1998 Spring) 70-72  
395-403.  
Journal code: 8208561. ISSN: 0273-2289.  
PUB. COUNTRY: United States  
DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
LANGUAGE: English  
FILE SEGMENT: Priority Journals  
ENTRY MONTH: 199808  
ENTRY DATE: Entered STN: 19980903  
Last Updated on STN: 19980903  
Entered Medline: 19980824

AB The saccharification of microcrystalline cellulose by reconstituted  
ternary mixtures of purified **cellulases** (one endoglucanase and  
two cellobiohydrolases) has been studied over the entire range of mixture  
compositions. Ternary plots are used to compare the performance of five  
synthetic mixtures drawn from the **cellulase** systems of  
**Acidothermus cellulolyticus**, *Trichoderma reesei*,  
*Thermomonospora fusca*, and *Thermotoga neapolitana*. Results reveal that at  
least one synthetic mixture utilizing enzymes from three different  
organisms delivers performance competitive with that of a "native" (i.e.,  
co-evolved) ternary system drawn exclusively from *T. reesei*. This  
heterologous system, consisting of the endoglucanase E1 from *A.*  
*cellulolyticus* and the **exoglucanases** CBHI from *T. reesei* and E3  
from *T. fusca*, is forgiving from the system-design point of view, in that  
it delivers high saccharification rates over a wide range of mixture  
compositions.

L13 ANSWER 2 OF 4 AGRICOLA DUPLICATE 2  
ACCESSION NUMBER: 96:12124 AGRICOLA  
DOCUMENT NUMBER: IND20499125  
TITLE: Quantitation of **Acidothermus**  
**cellulolyticus** E1 endoglucanase and  
*Thermomonospora fusca* E3 **exoglucanase** using  
enzyme-linked immunosorbent assay (ELISA).  
AUTHOR(S): Nieves, R.A.; Chou, Y.C.; Himmel, M.E.; Thomas, S.R.  
CORPORATE SOURCE: National Renewable Energy Laboratory, Golden, CO.  
AVAILABILITY: DNAL (QD415.A1J62)  
SOURCE: Applied biochemistry and biotechnology, Spring 1995.  
Vol. 51/52 p. 211-223  
Publisher: Totowa, N.J. : Humana Press.  
CODEN: ABIBDL; ISSN: 0273-2289  
NOTE: Paper presented at the Sixteenth Symposium on  
Biotechnology for Fuels and Chemicals, May 9-13, 1994,  
Gatlinburg, Tennessee.  
Includes references  
PUB. COUNTRY: New Jersey; United States  
DOCUMENT TYPE: Article  
FILE SEGMENT: U.S. Imprints not USDA, Experiment or Extension  
LANGUAGE: English

AB Two distinct quantitative indirect ELISAs were developed to determine the  
concentration of recombinant **cellulase** enzymes in culture  
filtrates. A monoclonal antibody (E1P7) was used as the primary antibody  
in developing an ELISA specific for **Acidothermus**

**cellulolyticus** E1 endoglucanase. Likewise, a polyclonal rabbit serum (Ab684) was used to develop an ELISA specific for *Thermomonospora fusca* E3 **exoglucanase**. Dose-response curves indicated a dynamic range for both assays between 0.01 and 0.08 micrograms/mL (1-8 ng/assay) when purified enzymes were used as standards. These assays have been used to estimate concentrations of secreted recombinant E1 and/or E3 in culture supernatants of *Streptomyces lividans* strain TK24 in which the corresponding genes have been cloned and expressed.

L13 ANSWER 3 OF 4 SCISEARCH COPYRIGHT 2003 ISI (R) DUPLICATE 3  
 ACCESSION NUMBER: 96:111961 SCISEARCH  
 THE GENUINE ARTICLE: BE51K  
 TITLE: SYNERGISM BETWEEN PURIFIED BACTERIAL AND FUNGAL  
**CELLULASES**  
 AUTHOR: BAKER J O (Reprint); ADNEY W S; THOMAS S R; NIEVES R A;  
 CHOU Y C; VINZANT T B; TUCKER M P; LAYMON R A; HIMMEL M E  
 CORPORATE SOURCE: NATL RENEWABLE ENERGY LAB, DIV ALTERNAT FUEL, APPLIED BIOL  
 SCI BRANCH, 1617 COLE BLVD, GOLDEN, CO, 80401 (Reprint)  
 COUNTRY OF AUTHOR: USA  
 SOURCE: ACS SYMPOSIUM SERIES, (1995) Vol. 618, pp. 113-141.  
 ISSN: 0097-6156.  
 DOCUMENT TYPE: General Review; Journal  
 LANGUAGE: ENGLISH  
 REFERENCE COUNT: 59

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

AB A standardized comparative study measured glucose release and synergistic effects in the solubilization of microcrystalline cellulose by binary mixtures of 11 fungal and bacterial **cellulases** (eight endoglucanases and three **exoglucanases**). Evaluation of 16 endo/exo pairs revealed that bacterial/fungal hybrid pairs are very effective in solubilizing microcrystalline cellulose. Of nine bacterial/fungal hybrid pairs studied, six were ranked among the nine most synergistic combinations, and six bacterial/fungal pairs were also among the top nine pairs in terms of soluble-sugar release. One hybrid pair (***Acidothermus cellulolyticus*** E1 and *Trichoderma reesei* CBH I) was ranked first in both synergism and sugar-release. In exo/exo synergism experiments, the performance of *Thermomonospora fusca* E(3) confirmed its mode of action as "CBH II-like" (i.e., E(3) is synergistic with *T. reesei* CBH I but not with *T. reesei* CBH II). Studies of endo/endo interactions suggested a possible means of categorizing endoglucanases in terms of substrate specificity.

L13 ANSWER 4 OF 4 CABA COPYRIGHT 2003 CABI DUPLICATE 4  
 ACCESSION NUMBER: 95:102060 CABA  
 DOCUMENT NUMBER: 950308088  
 TITLE: A new thermostable endoglucanase,  
***Acidothermus cellulolyticus*** E1.  
 Synergism with *Trichoderma reesei* CBH I and  
 comparison to *Thermomonospora fusca* E5  
 AUTHOR: Baker, J. O.; Adney, W. S.; Nieves, R. A.; Thomas,  
 S. R.; Wilson, D. B.; Himmel, M. E.  
 CORPORATE SOURCE: Alternative Fuels Division, National Renewable  
 Energy Laboratory, 1617 Cole Blvd., Golden, CO  
 80401, USA.  
 SOURCE: Applied Biochemistry and Biotechnology. Part A,  
 Enzyme Engineering and Biotechnology, (1994) Vol.  
 45/46, pp. 245-256. 19 ref.  
 Meeting Info.: Proceedings of the fifteenth  
 symposium on biotechnology for fuels and chemicals,  
 held at Colorado Springs, USA, 10-14 May 1993.  
 DOCUMENT TYPE: Conference Article; Journal  
 LANGUAGE: English  
 AB A new thermostable endoglucanase (***Acidothermus***  
***cellulolyticus*** E1) and another bacterial endoglucanase

(*Thermomonospora fusca* E5) each exhibit striking synergism with a fungal cellobiohydrolase (*Trichoderma reesei* CBH I [a cellulose 1,4- beta -cellulobiosidase]) in the saccharification of microcrystalline cellulose. In neither case did the endoglucanase:exoglucanase ratio that demonstrated maximum synergism coincide exactly with the ratio that actually released the maximum quantity of soluble sugar for a given total cellulase loading. The difference between the two ratios, after significant hydrolysis of the substrate, was considerably larger in the case of *A. cellulolyticus* E1. For both endoglucanase pairings with CBH I, the offset between the ratio for maximum synergism and the ratio for maximum soluble sugar production was found to be a function of digestion time.

=> d l14 ibib ab 1-17

L14 ANSWER 1 OF 17 MEDLINE DUPLICATE 1  
ACCESSION NUMBER: 2002354510 MEDLINE  
DOCUMENT NUMBER: 22092324 PubMed ID: 12096911  
TITLE: The crystal structure and catalytic mechanism of  
cellobiohydrolase CelS, the major enzymatic component of  
the Clostridium thermocellum Cellulosome.  
AUTHOR: Guimaraes Beatriz G; Souchon Helene; Lytle Betsy L; David  
Wu J H; Alzari Pedro M  
CORPORATE SOURCE: Unite de Biochimie Structurale, CNRS URA 2185, Institut  
Pasteur, 25 rue du Dr. Roux, 75724 Paris cedex 15, France.  
SOURCE: JOURNAL OF MOLECULAR BIOLOGY, (2002 Jul 12) 320 (3) 587-96.  
Journal code: 2985088R. ISSN: 0022-2836.  
PUB. COUNTRY: England: United Kingdom  
DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
LANGUAGE: English  
FILE SEGMENT: Priority Journals  
OTHER SOURCE: PDB-1L1Y; PDB-1L2A  
ENTRY MONTH: 200208  
ENTRY DATE: Entered STN: 20020707  
Last Updated on STN: 20020816  
Entered Medline: 20020815  
AB Cellobiohydrolase CelS plays an important role in the cellulosome, an  
active cellulase system produced by the thermophilic anaerobe  
Clostridium thermocellum. The structures of the catalytic domain of CelS  
in complex with substrate (cellohexaose) and product (cellobiose) were  
determined at 2.5 and 2.4 A resolution, respectively. The protein folds  
into an (alpha/alpha)(6) barrel with a tunnel-shaped substrate-binding  
region. The conformation of the loops defining the tunnel is intrinsically  
stable in the absence of substrate, suggesting a model to account for the  
processive mode of action of family 48  
cellobiohydrolases. Structural comparisons with other (alpha/alpha)(6)  
barrel glycosidases indicate that CelS and endoglucanase CelA, a  
sequence-unrelated family 8 glycosidase with a groove-shaped  
substrate-binding region, use the same catalytic machinery to hydrolyze  
the glycosidic linkage, despite a low sequence similarity and a different  
endo/exo mode of action. A remarkable feature of the mechanism is the  
absence, from CelS, of a carboxylic group acting as the base catalyst. The  
nearly identical arrangement of substrate and functionally important  
residues in the two active sites strongly suggests an evolutionary  
relationship between the cellobiohydrolase and endoglucanase families,  
which can therefore be classified into a new clan of glycoside hydrolases.  
(c) 2002 Elsevier Science Ltd.

L14 ANSWER 2 OF 17 SCISEARCH COPYRIGHT 2003 ISI (R) DUPLICATE 2  
ACCESSION NUMBER: 2003:30883 SCISEARCH  
THE GENUINE ARTICLE: 626VV  
TITLE: The major component of the cellulosomes of anaerobic fungi  
from the genus Piromyces is a family 48  
glycoside hydrolase  
AUTHOR: Steenbakkers P J M; Freelove A; van Cranenbroek B;  
Sweegers B M C; Harhangi H R; Vogels G D; Hazlewood G P;  
Gilbert H J; den Camp H J M O (Reprint)  
CORPORATE SOURCE: Univ Nijmegen, Fac Sci, Dept Microbiol, Toernooiveld 1,  
NL-6525 ED Nijmegen, Netherlands (Reprint); Univ Nijmegen,  
Fac Sci, Dept Microbiol, NL-6525 ED Nijmegen, Netherlands;  
Babraham Inst, Lab Mol Enzymol, Cambridge CB2 4AT,  
England; Univ Newcastle Upon Tyne, Dept Biol & Nutr Sci,  
Newcastle Upon Tyne NE1 7RU, Tyne & Wear, England  
COUNTRY OF AUTHOR: Netherlands; England  
SOURCE: DNA SEQUENCE, (JAN 2002) Vol. 13, No. 6, pp. 313-320.  
Publisher: TAYLOR & FRANCIS LTD, 4 PARK SQUARE, MILTON  
PARK,, ABINGDON OX14 4RN, OXON, ENGLAND.



ISSN: 1042-5179.  
DOCUMENT TYPE: Article; Journal  
LANGUAGE: English  
REFERENCE COUNT: 45

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

AB Sequencing of two cDNAs from the anaerobic fungi *Piromyces equi* and *Piromyces* sp. strain E2 revealed that they both encode a glycoside hydrolase (GH) **family 48 cellulase**, containing two C-terminal fungal dockerin domains. N-terminal sequencing of the major component of the *Piromyces* multi-enzyme **cellulase** /hemicellulase complex, termed the cellulosome, showed that these 80 kDa proteins corresponded to the GH **family 48** enzyme. These data show for the first time that GH **family 48 cellulases** are not confined to bacteria, and that bacterial and fungal cellulosomes share the same pivotal component.

L14 ANSWER 3 OF 17 MEDLINE DUPLICATE 3  
ACCESSION NUMBER: 2002161351 IN-PROCESS  
DOCUMENT NUMBER: 21889743 PubMed ID: 11893054  
TITLE: The *Clostridium cellulovorans* cellulosome: an enzyme complex with plant cell wall degrading activity.  
AUTHOR: Doi R H; Tamaru Y  
CORPORATE SOURCE: Section of Molecular & Cellular Biology, Division of Biological Sciences, University of California, Davis 95616, USA.. rhdoi@ucdavis.edu  
SOURCE: Chem Rec, (2001) 1 (1) 24-32. Ref: 33  
Journal code: 101085550. ISSN: 1527-8999.  
PUB. COUNTRY: United States  
DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
General Review; (REVIEW)  
(REVIEW, TUTORIAL)  
LANGUAGE: English  
FILE SEGMENT: IN-PROCESS; NONINDEXED; Priority Journals  
ENTRY DATE: Entered STN: 20020315  
Last Updated on STN: 20021211

AB Cellulose comprises a major portion of biomass on the earth, and the turnover of this material contributes to the CO<sub>2</sub> cycle. **Cellulases**, which play a major role in the turnover of cellulosic materials, have been found either as free enzymes that work synergistically, or as an enzyme complex called the cellulosome. This review summarizes some of the general properties of cellulosomes, and more specifically, the properties of the *Clostridium cellulovorans* cellulosome. The *C. cellulovorans* cellulosome is an extracellular enzyme complex with a molecular weight of about  $1 \times 10^6$ , and is comprised of at least ten subunits. The major subunit is the scaffolding protein CbpA, with a molecular weight of 189,000. This nonenzymatic subunit contains a cellulose binding domain (CBD) that binds the cellulosome to the substrate, nine conserved cohesins or enzyme binding domains, and four conserved surface layer homologous (SLH) domains. It is postulated that the SLH domains help to bind the cellulosome to the cell surface. The cellulosomal enzymes include **cellulases** (family 5 and 9 endoglucanases and a **family 48** exoglucanase), a mannanase, a xylanase, and a pectate lyase. The cellulosome is capable of converting *Arabidopsis* and tobacco plant cells to protoplasts. One of the endoglucanases, EngE, contains three tandemly repeated SLHs at its N-terminus, and therefore appears capable of binding to the scaffolding protein CbpA as well as to the cell surface. Cellulosomes can attack crystalline cellulose, but the free cellulosomal enzymes can attack only soluble and amorphous celluloses. Nine genes for the cellulosome are found in a gene cluster *cbpA-exgS-engH-engK-hbpA-engL-manA-engM-engN*. Other cellulosomal genes such as *engB*, *engE*, and *engY* are not linked to the major gene cluster or to each other. By determining the structure and function of the cellulosome, we hope to increase the efficiency of the cellulosome by genetic engineering techniques.

L14 ANSWER 4 OF 17

MEDLINE

DUPLICATE 4

ACCESSION NUMBER: 2001086784 MEDLINE  
 DOCUMENT NUMBER: 20558242 PubMed ID: 11106416  
 TITLE: Temporal secretion of a multicellulolytic system in Myxobacter sp. AL-1. Molecular cloning and heterologous expression of cel9 encoding a modular endocellulase clustered in an operon with cel48, an exocellobiohydrolase gene.  
 AUTHOR: Avitia C I; Castellanos-Juarez F X; Sanchez E; Tellez-Valencia A; Fajardo-Cavazos P; Nicholson W L; Pedraza-Reyes M  
 CORPORATE SOURCE: Institute of Investigation in Experimental Biology, Faculty of Chemistry, University of Guanajuato, Mexico.  
 SOURCE: EUROPEAN JOURNAL OF BIOCHEMISTRY, (2000 Dec) 267 (24) 7058-64.  
 Journal code: 0107600. ISSN: 0014-2956.  
 PUB. COUNTRY: GERMANY: Germany, Federal Republic of  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 ENTRY MONTH: 200101  
 ENTRY DATE: Entered STN: 20010322  
 Last Updated on STN: 20010322  
 Entered Medline: 20010118

AB The Gram-negative soil micro-organism Myxobacter sp. AL-1 possesses at least five extracellular **cellulases**, the production of which is regulated by the growth cycle. We cloned the complete gene for one of these **cellulases**, termed cel9, which encoded a 67-kDa modular family 9 endoglycohydrolase, which was produced during the stationary phase of growth and was strongly enhanced by avicel. The predicted product of cel9 matches the structural architecture of family 9 **cellulases** such as *Thermomonospora fusca* endo/exocellulase E4. Cel9 protein was synthesized in *Escherichia coli* from a multicopy plasmid and in *Bacillus subtilis* from the isopropyl thiogalactoside-inducible Pspac promoter and was purified from the culture medium. Thermal stability, optimum pH and temperature dependence of Cel9 were similar when expressed from either source, and were indistinguishable from related **cellulases** produced by thermophilic bacteria. Downstream from cel9 was found a partial ORF, designated cel48, the deduced product of which was highly similar to bacterial exocellobiohydrolases and processive endoglucanases belonging to **family 48** of the glycosyl hydrolases. The cel9 and cel48 genes appear to be arranged as part of an operon.

L14 ANSWER 5 OF 17

MEDLINE

ACCESSION NUMBER: 2000495112 MEDLINE  
 DOCUMENT NUMBER: 20389519 PubMed ID: 10931180  
 TITLE: Cloning, expression and characterization of a **family 48** exocellulase, Cel48A, from *Thermobifida fusca*.  
 AUTHOR: Irwin D C; Zhang S; Wilson D B  
 CORPORATE SOURCE: Department of Molecular Biology and Genetics, Cornell University, Ithaca, NY, USA.  
 SOURCE: EUROPEAN JOURNAL OF BIOCHEMISTRY, (2000 Aug) 267 (16) 4988-97.  
 Journal code: 0107600. ISSN: 0014-2956.  
 PUB. COUNTRY: GERMANY: Germany, Federal Republic of  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 ENTRY MONTH: 200010  
 ENTRY DATE: Entered STN: 20001027  
 Last Updated on STN: 20021210  
 Entered Medline: 20001018

AB The gene for a 104-kDa exocellulase, Cel48A, formerly E6, was cloned from

Thermobifida fusca into Escherichia coli and Streptomyces lividans. The DNA sequence revealed a type II cellulose-binding domain at the N-terminus, followed by a FNIII-like domain and ending with a glycosyl hydrolase **Family 48** catalytic domain. The enzyme and catalytic domain alone were each expressed in and purified from S. lividans and had very low catalytic activity on swollen cellulose, carboxymethyl cellulose, bacterial microcrystalline cellulose and filter paper. However, in synergistic assays on filter paper, the addition of Cel48A to a balanced mixture of T. fusca endocellulase and exocellulase increased the specific activity from 7.9 to 11.7 micromol cellobiose.min-1.mL-1, more than 15-fold higher than any single enzyme alone. Cel48A retained > 50% of its maximum activity from pH 5 to 9 and from 40 to 60 degrees C. Using SWISSMODEL, the amino-acid sequence of the Cel48Acd was modeled to the known structure of Clostridium cellulolyticum CelF. **Family 48** enzymes are remarkably homologous at 35% identity for all their catalytic domains and some of the properties of the 10 members are discussed.

L14 ANSWER 6 OF 17 MEDLINE  
 ACCESSION NUMBER: 2000437099 MEDLINE  
 DOCUMENT NUMBER: 20398177 PubMed ID: 10940036  
 TITLE: A scaffoldin of the Bacteroides cellulosolvens cellulosome that contains 11 type II cohesins.  
 AUTHOR: Ding S Y; Bayer E A; Steiner D; Shoham Y; Lamed R  
 CORPORATE SOURCE: Department of Biological Chemistry, The Weizmann Institute of Science, Rehovot, Israel.  
 SOURCE: JOURNAL OF BACTERIOLOGY, (2000 Sep) 182 (17) 4915-25.  
 Journal code: 2985120R. ISSN: 0021-9193.  
 PUB. COUNTRY: United States  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 OTHER SOURCE: GENBANK-AF224509  
 ENTRY MONTH: 200009  
 ENTRY DATE: Entered STN: 20000928  
 Last Updated on STN: 20000928  
 Entered Medline: 20000921

AB A cellulosomal scaffoldin gene, termed cipBc, was identified and sequenced from the mesophilic cellulolytic anaerobe Bacteroides cellulosolvens. The gene encodes a 2,292-residue polypeptide (excluding the signal sequence) with a calculated molecular weight of 242,437. CipBc contains an N-terminal signal peptide, 11 type II cohesin domains, an internal family III cellulose-binding domain (CBD), and a C-terminal dockerin domain. Its CBD belongs to family IIIb, like that of CipV from Acetivibrio cellulolyticus but unlike the family IIIa CBDs of other clostridial scaffoldins. In contrast to all other scaffoldins thus far described, CipBc lacks a hydrophilic domain or domain X of unknown function. The singularity of CipBc, however, lies in its numerous type II cohesin domains, all of which are very similar in sequence. One of the latter cohesin domains was expressed, and the expressed protein interacted selectively with cellulosomal enzymes, one of which was identified as a **family 48** glycosyl hydrolase on the basis of partial sequence alignment. By definition, the dockerins, carried by the cellulosomal enzymes of this species, would be considered to be type II. This is the first example of authentic type II cohesins that are confirmed components of a cellulosomal scaffoldin subunit rather than a cell surface anchoring component. The results attest to the emerging diversity of cellulosomes and their component sequences in nature.

L14 ANSWER 7 OF 17 CAPLUS COPYRIGHT 2003 ACS  
 ACCESSION NUMBER: 2000:327676 CAPLUS  
 TITLE: Attack of cellulose microcrystals by bacterial cellobiohydrolases.  
 AUTHOR(S): Gilkes, Neil R.; Imai, Tomoya; Kilburn, Doug; Warren,

Tony; Sugiyama, Junji  
 CORPORATE SOURCE: Department of Microbiology and Immunology, University  
 of British Columbia, Vancouver, BC, V6T 1Z3, Can.  
 SOURCE: Book of Abstracts, 219th ACS National Meeting, San  
 Francisco, CA, March 26-30, 2000 (2000), BTEC-057.  
 American Chemical Society: Washington, D. C.  
 CODEN: 69CLAC  
 DOCUMENT TYPE: Conference; Meeting Abstract  
 LANGUAGE: English  
 AB **Cellulase** systems from aerobic bacteria and fungi contain  
 similar cellobiohydrolases exoglucanases that remove cellobiosyl residues  
 from the ends of substrate mols. Related cellobiohydrolases from glucosyl  
 hydrolase family 6 are found in both bacteria and fungi and attack  
 substrates from the non-reducing end. Fungal cellobiohydrolases from  
 family 7 and bacterial cellobiohydrolases from **family 48**  
 belong to different enzyme families but both attack from the reducing end.  
 The directions of attack by Cellulomonas fimi cellobiohydrolases CbhA and  
 CbhB.

L14 ANSWER 8 OF 17 MEDLINE DUPLICATE 5  
 ACCESSION NUMBER: 1998429479 MEDLINE  
 DOCUMENT NUMBER: 98429479 PubMed ID: 9755156  
 TITLE: The crystal structure of the processive endocellulase Celf  
 of Clostridium cellulolyticum in complex with a  
 thiooligosaccharide inhibitor at 2.0 A resolution.  
 AUTHOR: Parsieglia G; Juy M; Reverbel-Leroy C; Tardif C; Belaich J  
 P; Driguez H; Haser R  
 CORPORATE SOURCE: Laboratoire d'Architecture et Fonction des Macromolécules  
 Biologiques, Institut de Biologie Structurale et  
 Microbiologie, Centre National de la Recherche  
 Scientifique, Marseille cedex 20, France.  
 SOURCE: EMBO JOURNAL, (1998 Oct 1) 17 (19) 5551-62.  
 Journal code: 8208664. ISSN: 0261-4189.  
 PUB. COUNTRY: ENGLAND: United Kingdom  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 OTHER SOURCE: PDB-1FCE  
 ENTRY MONTH: 199811  
 ENTRY DATE: Entered STN: 19990106  
 Last Updated on STN: 19990106  
 Entered Medline: 19981123  
 AB The mesophilic bacterium Clostridium cellulolyticum exports multienzyme  
 complexes called cellulosomes to digest cellulose. One of the three major  
 components of the cellulosome is the processive endocellulase Celf. The  
 crystal structure of the catalytic domain of Celf in complex with two  
 molecules of a thiooligosaccharide inhibitor was determined at 2.0 A  
 resolution. This is the first three-dimensional structure to be solved of  
 a member of the **family 48** glycosyl hydrolases. The  
 structure consists of an (alpha alpha)6-helix barrel with long loops on  
 the N-terminal side of the inner helices, which form a tunnel, and an open  
 cleft region covering one side of the barrel. One inhibitor molecule is  
 enclosed in the tunnel, the other exposed in the open cleft. The active  
 centre is located in a depression at the junction of the cleft and tunnel  
 regions. Glu55 is the proposed proton donor in the cleavage reaction,  
 while the corresponding base is proposed to be either Glu44 or Asp230. The  
 orientation of the reducing ends of the inhibitor molecules together with  
 the chain translation through the tunnel in the direction of the active  
 centre indicates that Celf cleaves processively cellobiose from the  
 reducing to the non-reducing end of the cellulose chain.

L14 ANSWER 9 OF 17 MEDLINE DUPLICATE 6  
 ACCESSION NUMBER: 1998361925 MEDLINE  
 DOCUMENT NUMBER: 98361925 PubMed ID: 9696784

TITLE: Cloning and DNA sequencing of the genes encoding Clostridium josui scaffolding protein CipA and cellulase CelD and identification of their gene products as major components of the cellulosome.

AUTHOR: Kakiuchi M; Isui A; Suzuki K; Fujino T; Fujino E; Kimura T; Karita S; Sakka K; Ohmiya K

CORPORATE SOURCE: Faculty of Bioresources, Mie University, Tsu 514, Japan.

SOURCE: JOURNAL OF BACTERIOLOGY, (1998 Aug) 180 (16) 4303-8.  
Journal code: 2985120R. ISSN: 0021-9193.

PUB. COUNTRY: United States

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

OTHER SOURCE: GENBANK-AB004845; GENBANK-AB011057

ENTRY MONTH: 199809

ENTRY DATE: Entered STN: 19980917  
Last Updated on STN: 20000303  
Entered Medline: 19980904

AB The Clostridium josui cipA and celD genes, encoding a scaffolding-like protein (CipA) and a putative cellulase (CelD), respectively, have been cloned and sequenced. CipA, with an estimated molecular weight of 120,227, consists of an N-terminal signal peptide, a cellulose-binding domain of family III, and six successive cohesin domains. The molecular architecture of C. josui CipA is similar to those of the scaffolding proteins reported so far, such as Clostridium thermocellum CipA, Clostridium cellulovorans CbpA, and Clostridium cellulolyticum CipC, but C. josui CipA is considerably smaller than the other scaffolding proteins. CelD consists of an N-terminal signal peptide, a family 48 catalytic domain of glycosyl hydrolase, and a dockerin domain. N-terminal amino acid sequence analysis of the C. josui cellulosomal proteins indicates that both CipA and CelD are major components of the cellulosome.

L14 ANSWER 10 OF 17 MEDLINE DUPLICATE 7

ACCESSION NUMBER: 1998154434 MEDLINE

DOCUMENT NUMBER: 98154434 PubMed ID: 9493383

TITLE: Properties and gene structure of a bifunctional cellulolytic enzyme (Cela) from the extreme thermophile 'Anaerocellum thermophilum' with separate glycosyl hydrolase family 9 and 48 catalytic domains.

AUTHOR: Zverlov V; Mahr S; Riedel K; Bronnenmeier K

CORPORATE SOURCE: Institute of Molecular Genetics, Russian Academy of Science, Moscow, Russia.

SOURCE: MICROBIOLOGY, (1998 Feb) 144 ( Pt 2) 457-65.  
Journal code: 9430468. ISSN: 1350-0872.

PUB. COUNTRY: ENGLAND: United Kingdom

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

OTHER SOURCE: GENBANK-Z86105

ENTRY MONTH: 199804

ENTRY DATE: Entered STN: 19980430  
Last Updated on STN: 19980430  
Entered Medline: 19980421

AB A large cellulolytic enzyme (Cela) with the ability to hydrolyse microcrystalline cellulose was isolated from the extremely thermophilic, cellulolytic bacterium 'Anaerocellum thermophilum'. Full-length Cela and a truncated enzyme species designated Cela' were purified to homogeneity from culture supernatants. Cela has an apparent molecular mass of 230 kDa. The enzyme exhibited significant activity towards Avicel and was most active towards soluble substrates such as CM-cellulose (CMC) and beta-glucan. Maximal activity was observed between pH values of 5 and 6 and temperatures of 95 degrees C (CM-cellulase) and 85 degrees C (Avicelase). Cellobiose, glucose and minor amounts of celotriose were

observed as end-products of Avicel degradation. The CelA-encoding gene was isolated from genomic DNA of 'A. thermophilum' by PCR and the nucleotide sequence was determined. The celA gene encodes a protein of 1711 amino acids (190 kDa) starting with the sequence found at the N-terminus of CelA purified from 'A. thermophilum'. Sequence analysis revealed a multidomain structure consisting of two distinct catalytic domains homologous to glycosyl hydrolase families 9 and 48 and three domains homologous to family III cellulose-binding domain linked by Pro-Thr-Ser-rich regions. The enzyme is most closely related to CelA of *Caldicellulosiruptor saccharolyticus* (sequence identities of 96 and 97% were found for the N- and C-terminal catalytic domains, respectively). Endoglucanase CelZ of *Clostridium stercoararium* shows 70.4% sequence identity to the N-terminal family 9 domain and exoglucanase Cely from the same organism has 69.2% amino acid identity with the C-terminal **family 48** domain. Consistent with this similarity on the primary structure level, the 90 kDa truncated derivative CelA' containing the N-terminal half of CelA exhibited endoglucanase activity and bound to microcrystalline cellulose. Due to the significantly enhanced Avicelase activity of full-length CelA, exoglucanase activity may be ascribed to the C-terminal **family 48** catalytic domain.

L14 ANSWER 11 OF 17 MEDLINE DUPLICATE 8  
 ACCESSION NUMBER: 1998437496 MEDLINE  
 DOCUMENT NUMBER: 98437496 PubMed ID: 9761829  
 TITLE: Crystallization of the catalytic domain of *Clostridium cellulolyticum* CeLF **cellulase** in the presence of a newly synthesized **cellulase** inhibitor.  
 AUTHOR: Reverbel-Leroy C; Parsieglia G; Moreau V; Juy M; Tardif C; Driguez H; Belaich J P; Haser R  
 CORPORATE SOURCE: Universite de Provence, Place Victor-Hugo, 13331 Marseille CEDEX 3, France.  
 SOURCE: ACTA CRYSTALLOGRAPHICA. SECTION D: BIOLOGICAL CRYSTALLOGRAPHY, (1998 Jan 1) 54 ( Pt 1) 114-8. Journal code: 9305878. ISSN: 0907-4449.  
 PUB. COUNTRY: Denmark  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 ENTRY MONTH: 199811  
 ENTRY DATE: Entered STN: 19990115  
 Last Updated on STN: 19990115  
 Entered Medline: 19981130

AB The catalytic domain of the CeIF processive endocellulase, a **family 48** glycosyl hydrolase from *Clostridium cellulolyticum* has been crystallized in the presence of a newly synthesized inhibitor (methyl 4-S-beta-cellobiosyl-4-thio-beta-cellobioside), by vapour diffusion, using PEG as a precipitant. The protein crystallizes in the orthorhombic P212121 space group and diffracts to a resolution of 2.0 Å. The unit-cell parameters are a = 61.4, b = 84.5, c = 121.9 Å.

L14 ANSWER 12 OF 17 MEDLINE DUPLICATE 9  
 ACCESSION NUMBER: 97237718 MEDLINE  
 DOCUMENT NUMBER: 97237718 PubMed ID: 9084173  
 TITLE: Structure of the *Clostridium stercoararium* gene cely encoding the exo-1,4-beta-glucanase Avicelase II.  
 AUTHOR: Bronnenmeier K; Kundt K; Riedel K; Schwarz W H; Staudenbauer W L  
 CORPORATE SOURCE: Institute for Microbiology, Technical University Munich, Federal Republic of Germany.  
 SOURCE: MICROBIOLOGY, (1997 Mar) 143 ( Pt 3) 891-8. Journal code: 9430468. ISSN: 1350-0872.  
 PUB. COUNTRY: ENGLAND: United Kingdom  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English  
FILE SEGMENT: Priority Journals  
OTHER SOURCE: GENBANK-D16670; GENBANK-L06942; GENBANK-L32742;  
GENBANK-L38827; GENBANK-U30321; GENBANK-Z69359  
ENTRY MONTH: 199705  
ENTRY DATE: Entered STN: 19970609  
Last Updated on STN: 19970609  
Entered Medline: 19970529

AB The nucleotide sequence of the cely gene coding for the thermostable exo-1,4-beta-glucanase Avicelase II of Clostridium stercoarium was determined. The gene consists of an ORF of 2742 bp which encodes a preprotein of 914 amino acids with a molecular mass of 103 kDa. The signal-peptide cleavage site was identified by comparison with the N-terminal amino acid sequence of Avicelase II purified from C stercoarium. The cely gene is located in close vicinity to the celZ gene coding for the endo-1,4-beta-glucanase Avicelase I. The Cely-encoding sequence was isolated from genomic DNA of C. stercoarium with the PCR technique. The recombinant enzyme produced in Escherichia coli as a LacZ'-Cely fusion protein could be purified using a simple two-step procedure. The properties of Cely proved to be consistent with those of Avicelase II purified from C. stercoarium. Sequence comparison revealed that Cely consists of an N-terminal catalytic domain flanked by a domain of 95 amino acids with unknown function joined to a type III cellulose-binding domain. The catalytic domain belongs to the recently proposed family L of cellulases (family 48 of glycosyl hydrolases).

L14 ANSWER 13 OF 17 SCISEARCH COPYRIGHT 2003 ISI (R)  
ACCESSION NUMBER: 97:742566 SCISEARCH  
THE GENUINE ARTICLE: XZ128  
TITLE: Attack of carboxymethylcellulose at opposite ends by two cellobiohydrolases from Cellulomonas fimi  
AUTHOR: Gilkes N R (Reprint); Kwan E; Kilburn D G; Miller R C; Warren R A J  
CORPORATE SOURCE: UNIV BRITISH COLUMBIA, DEPT MICROBIOL & IMMUNOL, 300-6174 UNIV BLVD, VANCOUVER, BC V6T 1Z3, CANADA (Reprint); UNIV BRITISH COLUMBIA, PROT ENGN NETWORK CTR EXCELLENCE, VANCOUVER, BC V6T 1Z3, CANADA  
COUNTRY OF AUTHOR: CANADA  
SOURCE: JOURNAL OF BIOTECHNOLOGY, (16 SEP 1997) Vol. 57, No. 1-3, pp. 83-90.  
Publisher: ELSEVIER SCIENCE BV, PO BOX 211, 1000 AE AMSTERDAM, NETHERLANDS.  
ISSN: 0168-1656.  
DOCUMENT TYPE: Article; Journal  
FILE SEGMENT: LIFE; AGRI  
LANGUAGE: English  
REFERENCE COUNT: 30

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

AB The patterns of reducing sugar release following sequential addition of two Cellulomonas fimi cellobiohydrolases to carboxymethylcellulose corroborate previous evidence for preferential attack of beta-1,4-glucans at opposite ends. In other experiments, sequential additions involving a single cellobiohydrolase and an Agrobacterium beta-glucosidase indicate that CbhA attacks preferentially from the non-reducing end, CbhB from the reducing end. We suggest that all aerobic bacteria and fungi involved in cellulose hydrolysis produce a similar pair of cellobiohydrolases: a family 6 cellobiohydrolase that attacks from the non-reducing end and a family 48 (bacterial) or family 7 (fungal) cellobiohydrolase that attacks from the reducing end. (C) 1997 Elsevier Science B.V.

L14 ANSWER 14 OF 17 MEDLINE DUPLICATE 10  
ACCESSION NUMBER: 97090419 MEDLINE

DOCUMENT NUMBER: 97090419 PubMed ID: 8936327  
 TITLE: Molecular study and overexpression of the Clostridium cellulolyticum celF **cellulase** gene in Escherichia coli.  
 AUTHOR: Reverbel-Leroy C; Belaich A; Bernadac A; Gaudin C; Belaich J P; Tardif C  
 CORPORATE SOURCE: Bioenergetique et Ingenierie des proteines, Centre National de la Recherche Scientifique, Marseille, France.  
 SOURCE: MICROBIOLOGY, (1996 Apr) 142 ( Pt 4) 1013-23.  
 Journal code: 9430468. ISSN: 1350-0872.  
 PUB. COUNTRY: ENGLAND: United Kingdom  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 OTHER SOURCE: GENBANK-U30321  
 ENTRY MONTH: 199612  
 ENTRY DATE: Entered STN: 19970128  
 Last Updated on STN: 19980206  
 Entered Medline: 19961231

AB The CelF-encoding sequence was isolated from Clostridium cellulolyticum genomic DNA using the inverse PCR technique. The gene lies between cipC (the gene encoding the cellulosome scaffolding protein) and celC (coding for the endoglucanase C) in the large cel cluster of this mesophilic cellulolytic Clostridium species. Comparisons between the deduced amino acid sequence of the mature CelF (693 amino acids, molecular mass 77626) and those of other beta-glycanases showed that this enzyme belongs to the recently proposed family L of **cellulases** (family 48 of glycosyl hydrolases). The protein was overproduced in Escherichia coli using the T7 expression system. It formed both cytoplasmic and periplasmic inclusion bodies when induction was performed at 37 degrees C. Surprisingly, the protein synthesized from the cytoplasmic production vector was degraded in the Ion protease-deficient strain BL21(DE3). The induction conditions were optimized with regard to the concentration of inductor, cell density, and temperature and time of induction in order to overproduce an active periplasmic protein (CelFp) which was both soluble and stable. It was collected using the osmotic shock method. The enzymic degradation of various cellulosic substrates by CelFp was studied. CelFp degraded swollen Avicel more efficiently than substituted soluble CM-cellulose or crystalline Avicel and was not active on xylan. Its activity is therefore quite different from that of endoglucanases, which are most active on CM-cellulose.

L14 ANSWER 15 OF 17 CAPLUS COPYRIGHT 2003 ACS

ACCESSION NUMBER: 1996:683734 CAPLUS  
 DOCUMENT NUMBER: 126:44206  
 TITLE: Similarities between bacterial and fungal **cellulase** systems  
 AUTHOR(S): Miller, R. C., Jr.; Gilkes, N. R.; Johnson, P.; Kilburn, D. G.; Kwan, E.; Meinke, A.; Schmuck, M.; Shen, H.; Tomme, P.; Warren, R. A. J.  
 CORPORATE SOURCE: Dept. Microbiol. Immunol., Univ. British Columbia, Vancouver, BC, V6T 1Z3, Can.  
 SOURCE: Biotechnology in the Pulp and Paper Industry: Recent Advances in Applied and Fundamental Research, Proceedings of the International Conference on Biotechnology in the Pulp and Paper Industry, 6th, Vienna, June 11-15, 1995 (1996), Meeting Date 1995, 531-536. Editor(s): Srebotnik, Ewald; Messner, Kurt. Facultas-Universitaetsverlag: Vienna, Austria.  
 CODEN: 63OUAN  
 DOCUMENT TYPE: Conference; General Review  
 LANGUAGE: English

AB A review with 28 refs. Six **cellulases** and two xylanases from the cellulolytic bacterium Cellulomonas fimi have now been characterized



in detail. All contain a catalytic domain and a cellulose-binding domain; in some, addnl. domains are present. The enzymes adopt extended conformations spanning ten or more cellobiosyl residues. This review is focused on two recently discovered *C. fimi* cellulases: CbhA and CbhB. The catalytic domain of CbhA is closely related to that of *Trichoderma reesei* CBH II in family 6 of the .beta.-1,4-glucanases. Biochem. analyses show that CbhA is an exocellobiohydrolase, like CBH II. By analogy to CBH II, CbhA attacks substrates from the non-reducing end. The second enzyme, CbhB, is also an exocellobiohydrolase. It is a member of a new family of bacterial .beta.-1,4-glucanases designated **family 48**. CbhB appears to attack substrates from the reducing end. We suggest that a pair of cellobiohydrolases, which attack opposite ends of cellulose mols., is a general feature of cellulase systems from aerobic bacteria and fungi.

L14 ANSWER 16 OF 17 SCISEARCH COPYRIGHT 2003 ISI (R)  
 ACCESSION NUMBER: 96:111964 SCISEARCH  
 THE GENUINE ARTICLE: BE51K  
 TITLE: CELLULOMONAS-FIMI CELLOBIOHYDROLASES  
 AUTHOR: SHEN H; MEINKE A; TOMME P; DAMUDE H G; KWAN E; KILBURN D G; MILLER R C; WARREN R A J; GILKES N R (Reprint)  
 CORPORATE SOURCE: UNIV BRITISH COLUMBIA, DEPT MICROBIOL & IMMUNOL, 300-6174 UNIV BLVD, VANCOUVER, BC V6T 1Z3, CANADA (Reprint); UNIV BRITISH COLUMBIA, DEPT MICROBIOL & IMMUNOL, VANCOUVER, BC V6T 1Z3, CANADA  
 COUNTRY OF AUTHOR: CANADA  
 SOURCE: ACS SYMPOSIUM SERIES, (1995) Vol. 618, pp. 174-196. ISSN: 0097-6156.  
 DOCUMENT TYPE: General Review; Journal  
 LANGUAGE: ENGLISH  
 REFERENCE COUNT: 66

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

AB The cellulolytic bacterium *Cellulomonas fimi* produces two cellobiohydrolases, CbhA and CbhB. These enzymes are major extracellular components during growth on cellulose. They correspond to previously identified cellulose-binding proteins, cbp95 and cbp120, respectively. Both comprise an N-terminal catalytic domain joined to three fibronectin type III modules and a C-terminal cellulose-binding domain. Amino acid sequence comparison shows that the CbhA catalytic domain is closely related to the catalytic domains of *Trichoderma reesei* CBH II and other fungal cellobiohydrolases in beta-1,4-glucanase family B. CbhB is a member of a new beta-1,4-glucanase family, designated family L (also called **family 48**). On the basis of data for CbhB, hydrolysis by family L enzymes proceeds with inversion of configuration at the anomeric carbon. CbhA and CbhB produce cellobiose from cellulose. Viscometric analysis of carboxymethylcellulose hydrolysis shows that both enzymes are predominantly exohydrolytic but Congo Red staining shows they also have weak endoglucanase activity. CbhB attacks cellohexaose from the reducing end. We suggest that *C. fimi* and *T. reesei* use similar strategies to hydrolyze cellulose because both appear to produce two types of cellobiohydrolase: one that attacks cellulose from the reducing end and another that attacks from the non-reducing end.

L14 ANSWER 17 OF 17 MEDLINE DUPLICATE 11  
 ACCESSION NUMBER: 96003898 MEDLINE  
 DOCUMENT NUMBER: 96003898 PubMed ID: 7575482  
 TITLE: Cellobiohydrolase B, a second exo-cellobiohydrolase from the cellulolytic bacterium *Cellulomonas fimi*.  
 AUTHOR: Shen H; Gilkes N R; Kilburn D G; Miller R C Jr; Warren R A  
 CORPORATE SOURCE: Department of Microbiology and Immunology, University of British Columbia, Vancouver, Canada.  
 SOURCE: BIOCHEMICAL JOURNAL, (1995 Oct 1) 311 ( Pt 1) 67-74. Journal code: 2984726R. ISSN: 0264-6021.  
 PUB. COUNTRY: ENGLAND: United Kingdom

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
LANGUAGE: English  
FILE SEGMENT: Priority Journals  
OTHER SOURCE: GENBANK-D16670; GENBANK-L06942; GENBANK-L32742;  
GENBANK-L38827; GENBANK-M87018  
ENTRY MONTH: 199511  
ENTRY DATE: Entered STN: 19951227  
Last Updated on STN: 19951227  
Entered Medline: 19951109

AB The gene cbhB from the cellulolytic bacterium *Cellulomonas fimi* encodes a polypeptide of 1090 amino acids. Cellobiohydrolase B (CbhB) is 1037 amino acids long, with a calculated molecular mass of 109765 Da. The enzyme comprises five domains: an N-terminal catalytic domain of 643 amino acids, three fibronectin type III repeats of 97 amino acids each, and a C-terminal cellulose-binding domain of 104 amino acids. The catalytic domain belongs to **family 48** of glycosyl hydrolases. CbhB has a very low activity on CM-cellulose. Viscometric analysis of CM-cellulose hydrolysis indicates that the enzyme is an exoglucanase. Cellobiose is the major product of hydrolysis of cellulose. In common with two other exoglycanases from *C. fimi*, CbhB has low but detectable endoglucanase activity. CbhB is the second exo-cellobiohydrolase found in *C. fimi*. Therefore, the **cellulase** system of *C. fimi* resembles those of fungi in comprising multiple endoglucanases and cellobiohydrolases.